

# ComASP<sup>®</sup> Colistin 0.25-16

System for colistin susceptibility testing with broth microdilution method, according to ISO 20776-1.

#### INTENDED PURPOSE

ComASP<sup>®</sup> Colistin 0.25-16 is an in vitro diagnostic product for antimicrobial susceptibility testing (AST) of clinical isolates based on growth of the test organism in the presence of various concentrations of an antimicrobial agent.

This ComASP<sup>®</sup> system is used for quantitative determination of the minimum inhibitory concentration (MIC) of colistin against the following non-fastidious organisms:

<u>Gram-negative bacteria</u> Enterobacterales Pseudomonas aeruginosa Acinetobacter spp.

#### DESCRIPTION

Clinicians are re-thinking to colistin as therapeutic option for the treatment of severe infections caused by multidrugresistant microorganisms, such as *Pseudomonas aeruginosa*, *Acinetobacter baumannii* and carbapenem-resistant *Enterobacterales*. However, colistin-resistant strains are worldwide disseminated leading to the need to determine the value of minimum inhibitory concentration (MIC) before the drug is prescribed. Although several commercial tests based on different techniques have been developed, broth microdilution (BMD) is considered the best method for performing colistin susceptibility testing so far.

ComASP<sup>®</sup> Colistin 0.25-16 is a 4-test panel containing the dried-up antibiotic in 7 two-fold dilutions:

#### Colistin concentration ranges 0.25 to 16 µg/ml.

The ComASP<sup>®</sup> system is a compact version of the broth microdilution (BMD) reference method allowing to perform the antimicrobial susceptibility testing of Colistin (polymyxin E) as recommended by international standards (i.e. CLSI, EUCAST, ISO) but in a simpler and less time-consuming way.

#### **KIT CONTENT**

- 4 Systems (panels) of ComASP<sup>®</sup> Colistin 0.25-16 (panels individually packed in foil with silica gel desiccant)
- 16 Tubes of Mueller Hinton II Broth (3.6 ml)
- Sealing Film
- Instructions Sheet and Test Results Form

#### MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as:

- loops pipettes physiological solutions swabs culture media incubator
- test tubes 0.5 McFarland turbidity standard quality control organisms

#### CONFIGURATION

Test	Colistin Concentration (µg/ml)							
Α	Growth	0.25	0.5	1	2	4	8	16
В	Growth	0.25	0.5	1	2	4	8	16
С	Growth	0.25	0.5	1	2	4	8	16
D	Growth	0.25	0.5	1	2	4	8	16

Growth indicates growth control: No antimicrobial agent in the well.

# PRINCIPLE OF THE METHOD

Four bacterial isolates may be tested on a single panel. All wells in a single row (test A, B, C or D) are rehydrated with a standardized microbial suspension. After incubation for 16-20 hours the result is read and interpreted.

#### SPECIMEN COLLECTION AND PREPARATION

ComASP<sup>®</sup> Colistin 0.25-16 is not for use directly with clinical or other specimens. The microorganism to be tested must first be isolated on a nonselective culture medium, such as blood agar or tryptic soy agar (TSA). In case of mixed culture, selected colonies should be purified by subculturing. Differential media harboring chromogenic or fluorogenic substrates should not be used for the subculture. It is recommended that cultures be no more than 24 hours old unless additional incubation is required to achieve sufficient growth.

#### **TEST PROCEDURE**

- 1. Take a panel from its envelope and leave it at room temperature for 10 min, DO NOT DISCARD THE ENVELOPE until all 4 tests have been carried out.
- 2. Prepare a suspension of the test organism using either the direct colony suspension or growth method.
- 3. Standardize the suspension to the density of a McFarland 0.5 standard.
- 4. Optimally within 15 min of preparation, dilute the adjusted suspension 1:20 in saline; this will be the **Solution A**.
- 5. Add 0.4 ml of Solution A to a tube of MH II Broth\* provided in the kit to obtain the **Solution B**.
- 6. Dispense 100  $\mu$ l of Solution B into each well in a row.
- 7. Cover the panel with the lid provided and incubate at  $36 \pm 2^{\circ}$ C for 16-20 hours in ambient air.
- \* Mueller Hinton II Broth (g/l): Beef Extract 3.0 g; Acid Hydrolysate of Casein 17.5 g; Starch 1.5 g; Distilled Water 1000 ml; pH 7.3 ± 0.1 (adjusted with appropriate salts to provide 20-25 mg/l of calcium and 10-12.5 mg/l of magnesium).

# **READING THE RESULTS**

At the end of the incubation period observe the growth in the wells and establish the MIC, i.e. the lowest concentration of antibiotic that inhibits visible growth.

The result can be read visually or automatically. Reading by the naked eye can be improved by use of bright indirect lighting against a dark background.

Growth appears as turbidity or as a button at the bottom of the well (compare with the amount of growth in the growth-control well).

The growth-control well should be evaluated first. Make sure it is positive for growth. If not, check the viability of the colonies picked and repeat the test using a new row in the same panel or a new panel and a microbial culture of recent growth.

Note the results on the Test Results Form included in the kit (copy as many form as necessary).

# INTERPRETATION OF THE RESULTS

Antimicrobial agant	Organism	MIC Criteria (µg/ml)		
Antimicrobial agent	Organishi	S≤	R>	
	Enterobacterales	2	2	
Colistin	Pseudomonas spp.	4	4	
	Acinetobacter spp.	2	2	

The MIC obtained should be interpreted according to current EUCAST interpretative criteria (see below).

**Disclaimer**: This breakpoint table might be out-of-date and does not replace EUCAST published guidelines, which always should be consulted before MIC categorization.

**NOTE**: Each individual panel is used for performing four tests. If less than 4 tests have been performed, use the film provided in the kit to seal the inoculated rows so to prevent any leakage of contaminated fluids. Then, return the panel into its own desiccant envelope and into the refrigerator (see STORAGE).

# **USER QUALITY CONTROL**

Quality control of ComASP<sup>®</sup> Colistin 0.25-16 is performed using the following reference strains:

1.	Escherichia coli	ATCC <sup>®</sup> 25922;	NCTC 12241
2.	Pseudomonas aeruginosa	ATCC <sup>®</sup> 27853;	NCTC 12903
3.	Escherichia coli	NCTC 13846	

Strain 1 and 2 are both susceptible to colistin and the acceptable **MIC QC range** is  $0.25-2 \mu g/ml$  for *E. coli* and  $0.5-4 \mu g/ml$  for *P. aeruginosa*. For *E. coli* NCTC 13846 (mcr-1 positive), which is instead resistant to colistin, the MIC target value is 4  $\mu g/ml$  and should only on occasion be 2 or 8  $\mu g/ml$ .

# PERFORMANCE CHARACTERISTICS

#### Accuracy

Accuracy of ComASP<sup>®</sup> Colistin 0.25-16 was determined by evaluating the agreement of the AST system result with the result generated for the same isolate with the broth microdilution (BMD) reference method. To assess accuracy, Essential Agreement (EA) and bias of the method were calculated. EA occurs when the MIC of the ComASP<sup>®</sup> system and the reference method agree exactly or is within  $\pm 1$  dilution of each other. Bias of the method is the evaluation of test device results to determine whether the results that differ from the reference method are significantly skewed or predominantly in one direction. A total of 300 clinical isolates were tested by three operators. The following table summarizes performance data from these studies.

Antimicrobial agent	Organism	Ν	%EA	%Bias
Colistin	Enterobacterales	174	96.6	13.5
	Pseudomonas aeruginosa	69	100	
	Acinetobacter spp.	57	98.2	
	TOTAL	300	97.7	

N, Number of isolates

EA, Essential Agreement

# Reproducibility

99.3% of ComASP<sup>®</sup> Colistin 0.25-16 results (3 *E. coli,* 1 *K. pneumoniae,* 1 *E. aerogenes,* 2 *A. baumannii* and 3 *P. aeruginosa* tested in triplicate by 3 operators on 3 days) were within a doubling dilution of reference microdilution results.

# Repeatability

100% of ComASP<sup>®</sup> Colistin 0.25-16 results (3 *E. coli,* 1 *K. pneumoniae,* 1 *E. aerogenes,* 2 *A. baumannii* and 3 *P. aeruginosa* tested in triplicate) were within a doubling dilution of reference microdilution results.

# LIMITATIONS

Invalid results can be caused by poor specimen quality, improper sample collection, improper transportation, improper laboratory processing, or a limitation of the testing technology. The operator should understand the principles of the procedures, including its performance limitations, in advance of operation to avoid potential mistakes.

# WARNINGS AND PRECAUTIONS

- 1) For in vitro diagnostic use (IVD) only.
- 2) For laboratory professional use only.
- 3) Operators must be trained and have certain experience. Please read the instructions carefully before using the kit. Reliability of assay results cannot be guaranteed if there are any deviations from the instructions in this document.
- 4) Do not use if a panel, tube or packaging appears to be damaged.
- 5) Follow standard precautions. All patient specimens should be considered potentially infectious and handled accordingly.
- 6) Handle all specimens as if infectious using safe laboratory procedures. Dispose of hazardous or biologically contaminated materials according to the practices of your institution.
- 7) Avoid cross-contamination of samples by using disposable tips and changing them after each sample.
- 8) Do not mix reagents of different batches. Please use the kit within the validity period.
- 9) Do not eat, drink, smoke, apply cosmetics or handle contact lenses in areas where reagents and human specimens are handled.
- 10) Results should be interpreted by a trained professional in conjunction with the patient's history and clinical signs and symptoms, and epidemiological risk factors.
- 11) Ensure laboratory equipment is calibrated and maintained in accordance with the laboratory's procedure.
- 12) When test results are transmitted from the laboratory to an informatics centre, attention has to be done to avoid erroneous data transfer.

# **STORAGE**

Store ComASP<sup>®</sup> Colistin 0.25-16 at 2-8°C in the original packaging. Once an envelope is opened the panel shall be used within 7 days and stored at 2-8°C. Do not use the panels beyond the expiry date indicated on the label. Eliminate without using if there are signs of deterioration.

# DISPOSAL OF USED MATERIAL

After use, ComASP<sup>®</sup> Colistin 0.25-16 and material that has come into contact with the sample must be decontaminated and disposed of in accordance with guidelines used in the laboratory for decontamination and disposal of potentially infected material.

# SUGGESTIONS FOR TROUBLESHOOTING

For out-of-range QC, first repeat the test with a pure culture or a freshly subcultured QC strain. If the issue is unresolved, follow this guidance for additional suggestions for troubleshooting out-of-range QC results and unusual clinical isolates results.

Observation	Probable Cause	Comments/Suggested Actions
MIC too low	Inoculum too light	Repeat using McFarland 0.5 turbidity
MIC too high	Inoculum too heavy	standard or standardizing device. Check expiration date and proper storage if using barium sulfate or latex standards. Check steps in inoculum preparation and incubation procedure. Perform colony count check of growth control well immediately after inoculation and before incubation ( <i>E.</i> <i>coli</i> ATCC <sup>®</sup> 25922 closely approximates 5 x 10 <sup>5</sup> CFU/ml)
MIC too high	Antimicrobial agent is degrading	Use alternative lot. Check STORAGE and package integrity
Skipped wells	Contamination. Inadequate mixing of inoculum or improper inoculation of panel	Repeat QC test

In case of other malfunctions or defects, contact immediately Liofilchem (\*) or the local representative.

In case of incident associated with the device, notify immediately Liofilchem (\*) or its local representative and the National Competent Authority.

\*Please login to <u>https://www.liofilchemstore.it/login.php</u> (user ID and password required) and click on Complaint.

# REFERENCES

- 1. Carretto E. *et al.* Clinical validation of the SensiTest<sup>™</sup> Colistin, a broth microdilution based method to evaluate colistin MICs. J Clin Microbiol. 2018;17. pii: JCM.01523-17. http://jcm.asm.org/content/early/2018/01/11/JCM.01523-17
- 2. CLSI. Performance Standards for Antimicrobial Susceptibility Testing. 31st ed. CLSI Supplement M100S. Wayne, PA: Clinical and Laboratory Standards Institute; 2021.
- 3. CLSI. Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. 11th ed. CLSI standard M07. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.
- 4. CLSI. Verification of Commercial Microbial Identification and Antimicrobial Susceptibility Testing Systems. 1st ed. CLSI guideline M52. Wayne, PA: Clinical and Laboratory Standards Institute; 2015.
- 5. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 11.0, 2021. http://www.eucast.org.
- 6. The European Committee on Antimicrobial Susceptibility Testing. Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST. Version 11.0, 2021. http://www.eucast.org.
- 7. ISO 20776-1:2019. Susceptibility testing of infectious agents and evaluation of performance of antimicrobial susceptibility test devices -- Part 1: Broth micro-dilution reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases.
- 8. Erlangga Y *et al*. The accuracy of four commercial broth microdilution tests in the determination of the minimum inhibitory concentration of colistin. Annals of Clinical Microbiology and Antimicrobials volume 19, Article number: 42 (2020) <u>https://doi.org/10.1186/s12941-020-00383-x</u>
- Sekyere JO et al. Comparative Evaluation of CHROMagar COL-APSE, MicroScan Walkaway, ComASP Colistin, and Colistin MAC Test in Detecting Colistin-resistant Gram-Negative Bacteria. Scientific Reports volume 10, Article number: 6221 (2020) <u>https://www.nature.com/articles/s41598-020-63267-2</u>
- 10. Galani I *et al.* Evaluation of ComASP<sup>™</sup> Colistin (formerly SensiTest<sup>™</sup> Colistin), a commercial broth microdilution-based method to evaluate the colistin minimum inhibitory concentration for carbapenem-resistant Klebsiella pneumoniae isolates. J Glob Antimicrobial Resistance. 2018; 15:123-126. DOI: <u>10.1016/j.jgar.2018.07.006</u>

A Summary of Safety and Performance (SSP) will be available on Eudamed (subject to Eudamed availability). This summary is also available on request at <u>liofilchem@liofilchem.com</u>

Product	Packaging	Ref.
ComASP <sup>®</sup> Colistin 0.25-16	4x4 tests	75001

# Table of Symbols

IVD	In Vitro Diagnostic Medical Device
REF	Catalogue number
LOT	Batch code
$\bigotimes$	Do not reuse
	Fragile, handle with care
<b>CE</b> <sub>0123</sub>	Identification number of notified body
	Manufacturer
X	Use by
Σ	Contains sufficient for <n> tests</n>
ĺ	Consult instructions for use
X	Temperature limits

# **Revision History**

Revision	Release Date	Change Summary
3.1	04 Sep 2018	Added schematic representation of test procedure
4	29 Sep 2023	Updated layout and content in compliance with IVDR 2017/746

**Note:** Minor typographical, grammar, and formatting changes are not included in revision history; Revision numbers are incremented by decimals (e.g. version 1.1 to v 1.2) when there are minor modifications that do not affect requirements or procedures.

This document is also available from the online Support Center: **liofilchem.com/ifu-sds** 

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EUCAST stands for European Committee on Antimicrobial Susceptibility Testing. These data have been made available at no cost by EUCAST and can be accessed freely on the EUCAST website: www.eucast.org. EUCAST recommendations are frequently updated and the latest versions are available at www.eucast.org.

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# TEST RESULTS FORM ComASP<sup>®</sup> Colistin 0.25-16

Patient data
Name
Surname
Age
Gender
Specimen
Notes

Indicate with the sign + the microbial growth.

Indicate with the sign – the absence of microbial growth.

Read the MIC and interpret the result according to the current EUCAST interpretive criteria.

TEST	COLISTIN CONCENTRATION (µg/ml)					MIC VALUE AND INTERPRETATION			
	Growth	0.25	0.5	1	2	4	8	16	
P	Growth	0.25	0.5	1	2	4	8	16	
D									
6	Growth	0.25	0.5	1	2	4	8	16	
D	Growth	0.25	0.5	1	2	4	8	16	

Use a distinct test results form for each sample under investigation. A horizontal line should be drawn on the empty spaces related to an unused test, if any.

Date of Test	Operator